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Esterase-Sensitive Prodrugs with Tunable Release Rates and Direct Generation of Hydrogen Sulfide**

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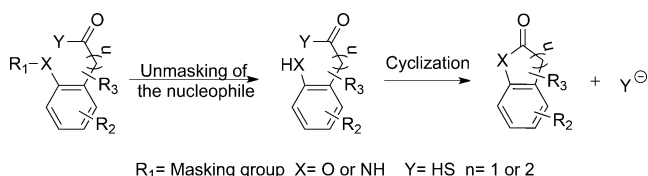
Abstract: Prodrugs that release hydrogen sulfide upon esterase-mediated cleavage of an ester group followed by lactonization are described herein. By modifying the ester group and thus its susceptibility to esterase, and structural features critical to the lactonization rate, H₂S release rates can be tuned. Such prodrugs directly release hydrogen sulfide without the involvement of perthiol species, which are commonly encountered with existing H₂S donors. Additionally, such prodrugs can easily be conjugated to another non-steroidal anti-inflammatory agent, leading to easy synthesis of hybrid prodrugs. As a biological validation of the H₂S prodrugs, the anti-inflammatory effects of one such prodrug were examined by studying its ability to inhibit LPS-induced TNF- α production in RAW 264.7 cells. This type of H₂S prodrugs shows great potential as both research tools and therapeutic agents.

H₂S is an endogenously produced signaling molecule in mammals and is critical to human health and disease.^[1] Many studies have also shown the various therapeutic effects of H₂S, which include protection against myocardial ischemia injury, cytoprotection against oxidative stress, mediation of neurotransmission, inhibition of insulin signaling, regulation of inflammation and dilation of blood vessel.^[1a,b,d,e,2]

In studying the physiological and pathological properties of H₂S, H₂S gas or inorganic sulfide salts such as Na₂S and NaHS have been widely used. However, the uncontrollable release of H₂S from sulfide salts and the toxic effects of excessive H₂S limit their potential as possible therapeutic agents.^[3] Moreover, inorganic sulfide salts could not mimic the slow and continuous H₂S production in the biological system, which further limits their usage. Therefore, new H₂S releasing agents (H₂S donors) are of great clinical and research interests.^[3,4] Currently, there are seven types of H₂S donors that have been reported:^[5] 1) garlic and related sulfur compounds;^[6] 2) Lawesson's reagent and analogs (GYY4137);^[7] 3) 1,2-dithiole-3-thiones (DTTs) and hybrids of H₂S and non-steroidal anti-inflammatory drugs;^[8] 4) thiol-activated H₂S donors;^[9] 5) photo-induced H₂S donors;^[10] 6) thiol amino acid;^[11] and 7) polysulfide (SG-1002).^[12] There are some obvious limitations among these H₂S donors. First of all, the release rates of H₂S from most

donors such as GYY4137 and DTTs are hard to control, and the effect of their byproducts associated with H₂S release is unclear. Furthermore, existing H₂S donor systems lack well defined negative controls. Some H₂S donors, such as thiol-activated H₂S donors would generate the perthiol intermediates, which may also have certain physiological effect, making it hard to deconvolute the experimental results. Almost all existing donors release precursors of H₂S. Thus the correlation of H₂S release and the amount of donor added is difficult. H₂S donors that can mimic endogenous H₂S production through a single enzymatic step are currently not available. Recently, Moore et al.^[13] tested the effect of GYY4137 on the release of pro- and anti-inflammatory mediators in lipopolysaccharide (LPS)-treated murine RAW264.7 macrophages, and found that the effects of H₂S on inflammatory processes are complex and dependent not only on H₂S donor concentration but also on the rate of H₂S generation. Therefore, there is a need for new H₂S prodrugs, which directly generate H₂S with well-defined and tunable release rates that are not directly affected by the redox balance of the cellular environment and the presence of other thiol species. Such H₂S donors/prodrugs will be very important research tools in delineating the functions of H₂S and lay a foundation for the future development of H₂S-based therapeutics.

Our lab has long-standing interests in prodrugs based on intramolecular cyclizations.^[14] We took advantage of one such lactonization prodrug systems (Scheme 1), and designed



Scheme 1. The general concept of cyclization-driven prodrugs of hydrogen sulfide.

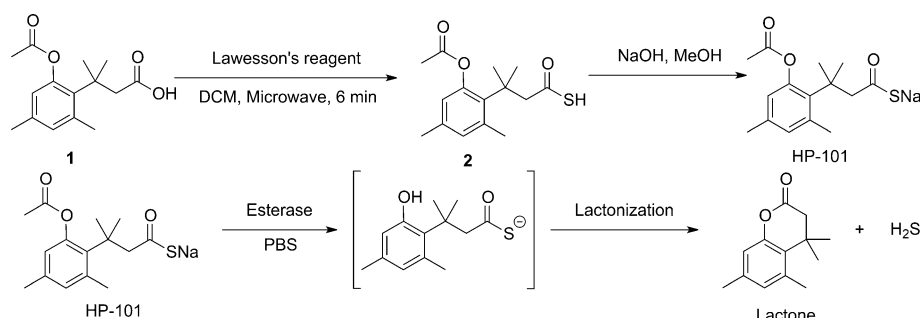
esterase-sensitive prodrugs of H₂S. Specifically, the nucleophilic hydroxy or amino group can be masked as an ester or amide, and the drug, H₂S, can be conjugated to the carbonyl carbon in the form of a thioacid. After hydrolysis of the masking group, the nucleophile can attack the carbonyl group and undergo a lactonization reaction, and thus release hydrogen sulfide.

To test the idea, we first synthesized prodrug BW-HP-101, which uses a "trimethyl lock" and thus stereochemical control to facilitate lactonization.^[14a,15] The synthesis of BW-HP-101 was accomplished by treating compound **1**,^[16] with Lawesson's reagent under microwave conditions,^[17] followed

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[**] Patent pending covering hydrogen sulfide precursors and drug conjugates thereof.

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Scheme 2. Synthesis of “trimethyl lock”-based H₂S prodrug HP-101 and the mechanism of esterase-triggered H₂S release.

by reaction with one equivalent of sodium hydroxide (Scheme 2).

The novel BW-HP-101 is a very stable white and odorless solid, and has very good water solubility, allowing 10 mM stock solutions to be prepared in aqueous buffer. The prodrug showed no obvious decomposition during storage at room temperature for 3 days and at -20°C for 3 month.

To examine the feasibility of the concept described in Schemes 1 and 2, we studied whether esterase can catalyze H₂S release from BW-HP-101 by using a H₂S-selective microelectrode, and found time-dependent H₂S release in the presence of porcine liver esterase (PLE) (Figure 1) with

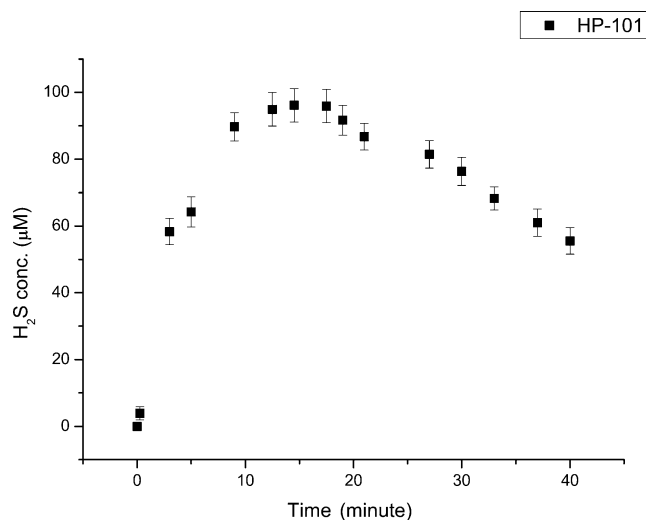


Figure 1. Hydrogen sulfide generation from BW-HP-101. 200 μM prodrug in PBS (1 % DMSO) at 37°C with 1 unit/mL of PLE ($p=0.95$, $n=3$).

a peak concentration at about 15 min. H₂S release was further confirmed by using a well-known hydrogen sulfide fluorescent probe WSP-5^[18] (Figure 2). Strong fluorescence was detected when WSP-5 was incubated with the prodrug in the presence of PLE in phosphate buffer saline (PBS) or cell culture media containing fetal bovine serum (FBS). Such results indicate that BW-HP-101 indeed releases H₂S in PBS with esterase catalysis, or cell culture media containing FBS. In contrast, incubation in PBS alone did not lead to H₂S formation, indicating the chemical stability of the prodrug.

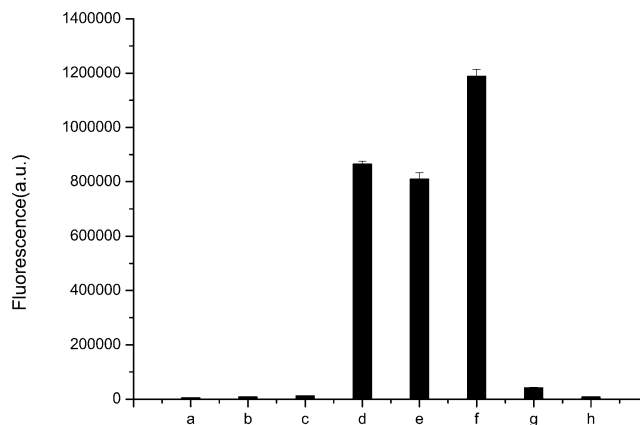
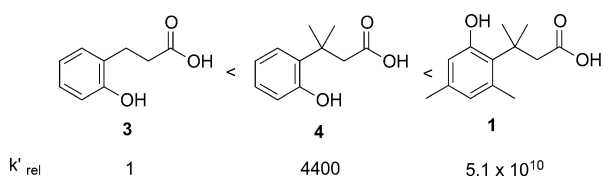


Figure 2. Qualitative detection of hydrogen sulfide release from BW-HP-101 by WSP-5. The concentration of WSP-5 is 50 μM , and the intensities of fluorescence were recorded after 5 min of incubation of WSP-5 with different substrates at room temperature. a) WSP-5 only in PBS; b) WSP-5 + 200 μM prodrug in PBS; c) WSP-5 + 200 μM prodrug in DMEM (Dulbecco's modified Eagle's medium; no FBS), no cells; d) WSP-5 + 200 μM prodrug in DMEM (with FBS) + cells; e) WSP-5 + 200 μM prodrug in DMEM (with FBS), no cells; f) WSP-5 + 200 μM prodrug + 1 unit/mL of esterase; g) WSP-5 + 200 μM GYY1437; h) WSP-5 + 1 unit/mL esterase.

ther studied the tunability of the release rates by varying the ester group and factors controlling the lactonization step. Variations of the ester group allows for tuning the rate of the unmasking step. For ester hydrolysis, it has been shown that increasing the size of the acyl moiety results in decreased catalytic hydrolysis rate.^[19] Thus we reasoned that modifying the acyl moiety should help tune the hydrolysis rates. The second direction in tuning H₂S releasing rates is based on controlling the lactonization rate by varying the number of methyl groups in the system. It is well known that the lactonization of compound **1** is much faster than that of *o*-hydroxydihydrocinnamic acids **3** and **4**, which lack pendant methyl groups (Scheme 3) and thus has decreased entropic control of the conformation favorable for lactonization.^[15b] Therefore, BW-HP-102, -103, and -104 were synthesized to tune the release rates. BW-HP-102 and BW-HP-104 contain a large acyl moiety cyclopropanecarbonyl ester and BW-HP-103 and BW-HP-104 lack two methyl groups on phenyl ring.

H₂S release from these prodrugs was studied (Figure 3). As designed, these prodrugs show very different H₂S release



Scheme 3. Relative rate constants for lactonization of various hydroxy-dihydrocinnamic acids.^[20]

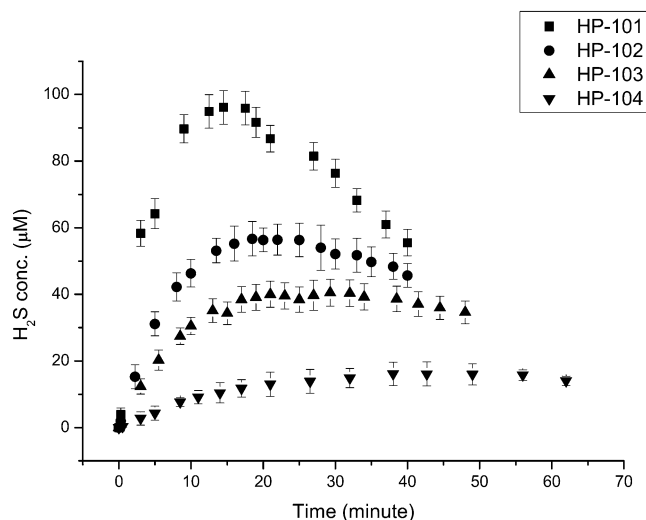
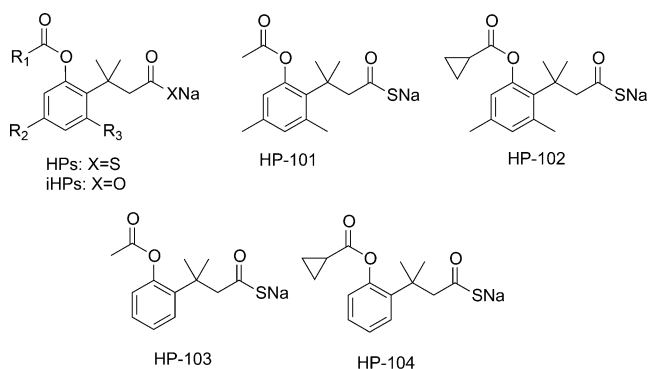


Figure 3. H_2S generation curves. 200 μM prodrugs in PBS (1% DMSO) with 1 unit/mL esterase at 37°C ($p=0.95$, $n=3$).

rates. For 200 μM of the prodrug in PBS at 37°C with 1 unit/mL PLE, the peak H_2S concentration for the fastest one, BW-HP-101, was about 95 μM at 15 min; and for the slowest one, BW-HP-104, it was about 13 μM at 43 min. Such results bring out an important issue, that is, the same concentration of the prodrug may mean very different effective H_2S concentrations, depending on the release rates. This issue is especially important in delivering a gaseous molecule because of its volatility nature and the lack of “accumulation”, which is in direct contrast to the delivery of non-volatile drugs. In the latter case, a slower release may only affect the onset time, but not the final concentration. With a gaseous molecule, release rates affect the onset time, the peak concentration, and the eventual concentration significantly. Thus when comparing the results using various H_2S prodrugs/donors, particular

attention needs to be paid to the release rate and effective concentration issue.

We also monitored the lactone product formation by HPLC (Table 1), and found $t_{1/2}$ ranging from 13 to 99 min for 200 μM prodrugs in the presence of PLE. Such results further demonstrated the concept of tuning the H_2S release rates.

Table 1: The half-lives of various prodrugs. 200 μM prodrugs in PBS with esterase 1 unit/mL at 37°C, $p=0.95$, $n=3$.

	BW-HP-101	BW-HP-102	BW-HP-103	BW-HP-104
$t_{1/2}$ [min]	13.0 ± 2.4	28.7 ± 1.5	44.5 ± 2.1	99.0 ± 8.9

The conjugation of two drugs with the same therapeutic indication, but different mechanisms, is attracting a great deal of attention in the hybrid drug field.^[21] Especially interesting is the idea of conjugating a non-steroidal anti-inflammatory drug (NSAID) to a H_2S donor,^[22] which has known anti-inflammatory effects. H_2S -NSAIDs have shown remarkable improvements in activity and tolerability as compared with the related parent compounds.^[21] However, few of the hydrogen sulfide prodrugs could be successfully applied to hybrid drug preparation for chemistry reasons, and existing H_2S -NSAIDs also suffer from the lack of control in H_2S release. Because most NSAIDs have a free carboxyl group, their conjugation to one of our prodrugs is not only easy, but also leads to a hybrid prodrug, which uses the same mechanism to release both drugs. This is very unique among all known H_2S hybrid drugs. Thus, we also synthesized BW-HP-105, which is formed by coupling the novel H_2S release system to naproxen (Figure 4). HPLC kinetic studies showed that BW-HP-105 could generate naproxen and H_2S by treatment with an esterase. Compared to other H_2S prodrugs with the same stereochemical control (BW-HP-101, 102), BW-HP-105 showed a slower hydrolysis rate as expected because of the larger masking group (naproxen as the acyl group). The H_2S -NSAID hybrid shown here is the first example of controllable H_2S release employing the same mechanism to “activate” both drugs, and will be a very useful research tool and potential therapeutic agent.

We next tested whether such compounds also produced H_2S -associated biological effects in vitro. For such studies, negative control compounds are always important. However, there are no good control compounds for existing H_2S prodrugs. To address this issue, we used the inactive oxyacid version of the prodrugs (iHPs, Figure 3) as the control compounds. Compared to thioacid prodrugs, iHPs have the same chemical structural frame except replacing sulfur with oxygen.

We first tested the cytotoxicity of these prodrugs on RAW264.7 macrophages. None showed any toxicity at 200 μM (see SI). We then examine the effect of the prodrugs on TNF- α production after co-treatment of the cell with the prodrugs and 1 $\mu g mL^{-1}$ LPS for 1 hour, using an ELISA kit (Figure 5). The results showed that only the prodrugs and GYY 4137 effectively inhibited TNF- α secretion, and Na_2S showed pro-inflammatory effect, which is similar to literature results.^[13] None of the iHPs showed the same effect, which clearly

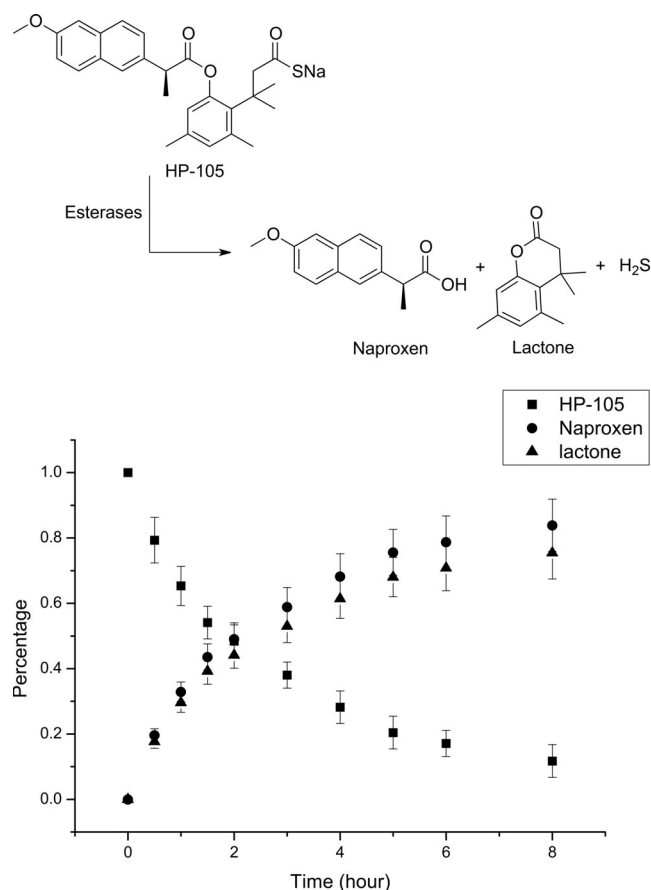


Figure 4. 200 μM HP-105 in PBS (1% DMSO) with esterase 20 unit/mL at 37°C ($p = 0.95$, $n = 3$).

demonstrated that the inhibition effect on TNF- α production came from the H_2S released from the respective prodrug.

In conclusion, we successfully investigated a new strategy of making H_2S prodrugs by using an esterase catalyzed lactonization prodrug system. Compared to existing H_2S prodrugs/donors, the new H_2S prodrugs described show several unique features. First of all, they have controlled H_2S release rates. This aspect seems to be the most challenging and important issue in the field of H_2S donors. Secondly, the trigger is an enzyme ubiquitous in the biological system.^[23] Thirdly, the prodrugs require a specific type of enzyme to trigger H_2S release, which afford the potential for controlled release at preferred sites. Fourthly, as research tools, the prodrugs described have well-defined negative controls. Fifthly, this strategy provides the first H_2S -NSAIDs hybrids with controllable release rates. We believe that these new H_2S prodrugs will be very useful research tools to others working in this field.

Acknowledgements

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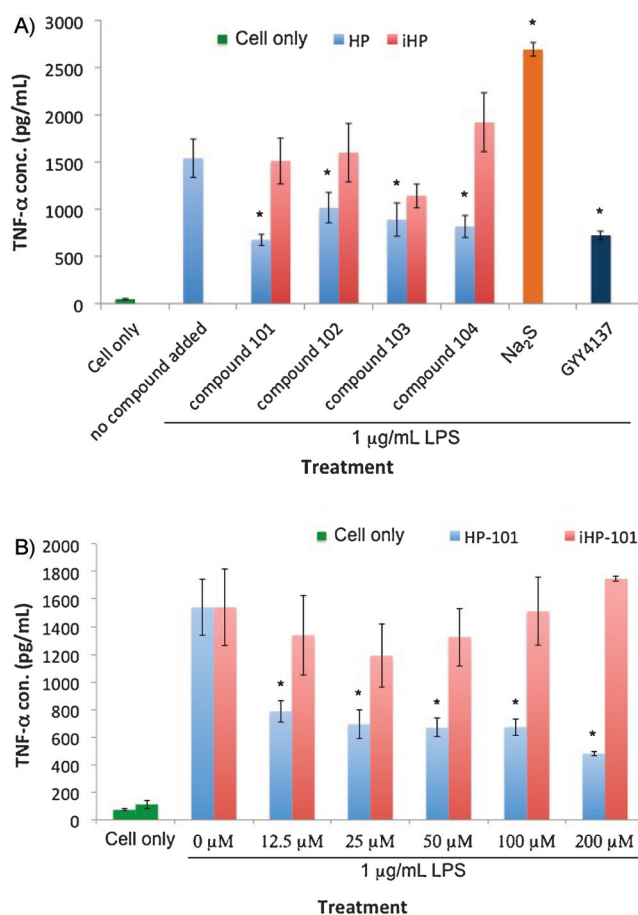


Figure 5. TNF- α concentrations of RAW 264.7 cell culture media after 1-hour co-treatment with H_2S prodrugs and LPS. A) Treatment with 50 μM HPs, iHPs Na_2S and GYY4137. B) Treatment with various concentrations of BW-HP-101 and iHP-101 ($n = 4$; $p < 0.05$).

Keywords: anti-inflammatory drugs · enzyme trigger · H_2S -NSAIDs · hydrogen sulfide prodrugs · lactonization

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